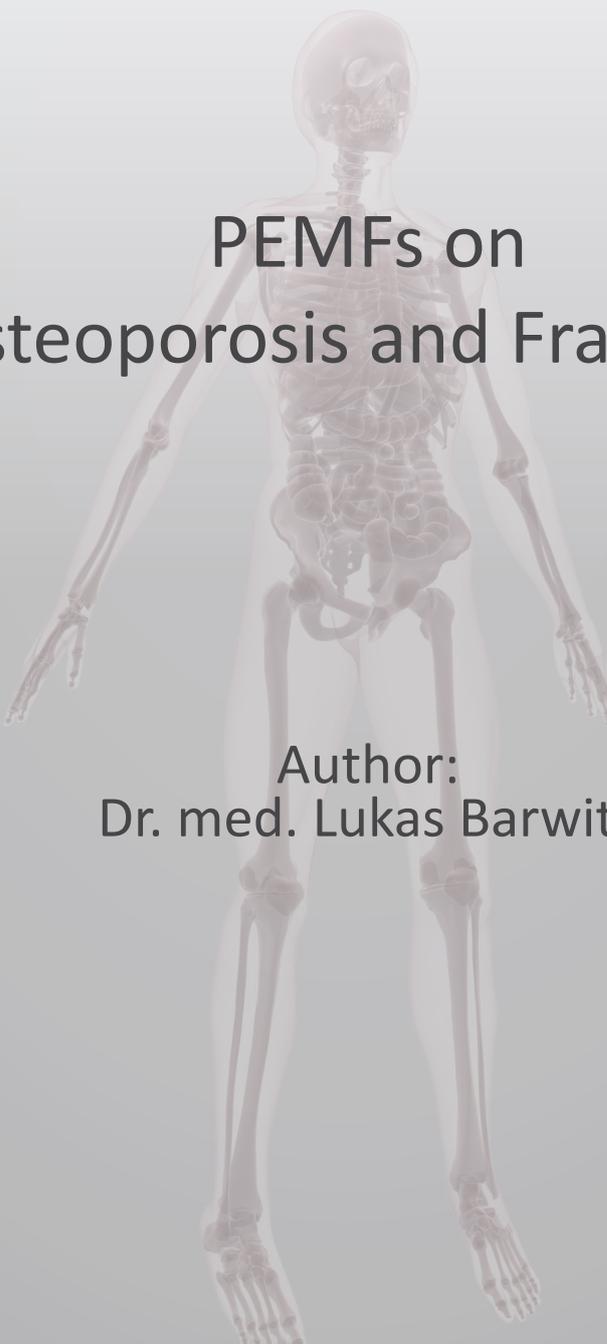


Bone Metabolism

A faint, semi-transparent illustration of a human skeleton is centered on the page, serving as a background for the text.

PEMFs on
Osteoporosis and Fractures

Author:
Dr. med. Lukas Barwitz

Bone metabolism is a continual cycle of bone growth and resorption that is carefully orchestrated by the dynamic relationship between osteoclasts, osteoblasts and an array of hormonal and regulatory influences. The relative levels of these signaling molecules dictate whether healthy, balanced bone metabolism ensues. Disturbances to this delicate equilibrium where resorption is greater than growth can weaken the skeletal architecture and put one at risk for the development of chronic and debilitating diseases such as Osteoporosis.

Osteoporosis

Osteoporosis is a disease where bone mass atrophy and the destruction of the microarchitecture of bone tissue increases the risk of fractures. It becomes more common with age and is the most common reason for a broken bone among the elderly. About 15% in their 50s and 70% of those over 80 are affected; it is more common in women than men.

Bones that commonly break include the vertebrae, the bones of the forearm, and the hip. Until a broken bone occurs there are typically no symptoms. Bones may weaken to such a degree that a fracture may occur with minor stress or even spontaneously. Chronic pain and a decreased ability to carry out normal activities may follow a fracture.

Bone loss increases after menopause due to lower levels of estrogen. Osteoporosis may also occur due to a number of diseases or treatments, including alcoholism, anorexia hyperthyroidism or kidney disease.

Certain medications increase the rate of bone loss, including some antiseizure medications, chemotherapy, proton pump inhibitors, selective serotonin reuptake inhibitors, and glucocorticosteroids. Smoking and too little exercise are also risk factors. Osteoporosis is defined as a bone density of 2.5 standard deviations below that of a young adult. This is typically measured by dual-energy X-ray absorptiometry.

Efforts to prevent broken bones in those with osteoporosis include a good diet, exercise, and fall prevention. Lifestyle changes such as stopping smoking and not drinking alcohol may help. Studies showed that Biphosphonate medications are useful in those with previous broken bones due to osteoporosis. In those with osteoporosis but no previous broken bones, they are less effective. A number of other medications may also be useful.

History of PEMFs on bone repair

In 1892, Wolf indicated that mechanical stress determines bone growth and remodelingⁱ. In 1953, Yasuda revealed that bending the long tubular bone is related with the development of electric currents and this instance is defined as piezoelectric phenomenonⁱⁱ. Since then, the theory that electrical stimulation is the path for bone formation in response to applied load has been gradually recognized, and various devices have been developed to produce

electrical stimulation for promoting the healing of bone fracture. In 1978, Bassett first applied noninvasive PEMFs to treat delayed union or non-union fractures and have achieved good clinical effectⁱⁱⁱ. Shortly thereafter, PEMFs were approved as a safe and effective method for treating delayed union or non-union fractures by the US Food and Drug Administration (FDA)^{iv,v}. Inductive coupling is the rationale for the application of PEMFs^{vi}. PEMF devices consist of a wire coil wherein a current passes and a pulsed magnetic field is generated. The pulsed magnetic field, in turn, induces a time-varying secondary electrical field within the bone. The secondary electrical field is dependent on the characteristics of the applied pulsed magnetic field and the tissue properties. Magnetic fields of 0.1–20 G are usually applied to produce electrical fields, ranging from 1 mV/cm to 100 mV/cm in the bone^{vii}. Through the PEMF device, a time-varying electrical field is produced to simulate the normal response of bone cells physiologically to the applied mechanical stress^{viii}, and the subsequent enhanced growth and remodeling bioeffects on the bone are initiated by the time-varying electrical field.

PEMFs on Osteoporosis and Fractures

In numerous studies PEMF could demonstrate its positive effects on bone metabolism and in the healing of fractures by stimulating osteoblasts, calcium uptake and its mineralization in vitro and in vivo and is therefore a suitable addition for the treatment of osteoporosis and fractures^{ix,x,xi,xii,xiii,xiv,xv,xvi,xvii,xviii,xix}.

What exactly happens on a cellular level?

Recently, considerable research progresses have been made in exploring the underlying cellular and subcellular mechanisms of PEMFs promotion effect in bone repair. Several key signaling pathways during the osteogenesis and angiogenesis which are two essential aspects for bone repair, were revealed by various studies when the bone was exposed to PEMF including Ca²⁺, Wnt/ β -catenin, mitogen-activated protein kinase (MAPK), fibroblast growth factor (FGF) and vascular endothelial growth factor (VEGF), transforming growth factor (TGF)- β /bone morphogenetic proteins (BMP), insulin-like growth factor (IGF), Notch, and cAMP/protein kinase A (PKA)^{xx}.

For those still interested in the very detailed signaling pathways during the osteogenesis and angiogenesis that PEMF takes influence on I recommend **Yuan J et al. 2018. Underlying Signaling Pathways and Therapeutic Applications of Pulsed Electromagnetic Fields in Bone Repair^{xx}** and quote:

Ca²⁺ Signaling

Intracellular Ca²⁺ is generally considered as one of the main actors to translate the PEMF signal into a biological signal. Many studies revealed that PEMF signal passes through the cell membrane to set up a time-varying electrical field within the cytosol; this electrical field subsequently induces the release of intracellular Ca²⁺, leading to increases in cytosolic calcium, activated calmodulin and the enhancement of bone cell viability^{viii,xxi,xxii}.

Voltage-gated Ca channels (VGCCs), especially the L type, play a pivotal role in intracellular Ca²⁺ release. PEMF exposure significantly elevated the expression levels of VGCCs in mesenchymal stem cells (MSCs) during osteogenesis^{xxiii,xxiv}. PEMF-initiated Ca²⁺ signaling strikingly accelerates the osteogenic differentiation of MSCs as represented by the upregulated osteogenic markers, such as collagen I and ALP, and the increased deposition of extracellular calcium^{xviii}. Accumulated studies indicated that increased intracellular Ca²⁺ caused by PEMF stimulation leads to increased nitric oxide levels, which in turn increases the synthesis level of cGMP and the subsequent activation of protein kinase G. Through the Ca²⁺/nitric oxide/ cGMP/protein kinase G pathway, PEMFs promote osteoblast differentiation and maturation, exert their therapeutic effect on bone repair, and remarkably reduce the pain of patients by modulating the release of inflammatory cytokines, such as interleukin-1 beta (IL-1β)^{xvi,xxv,xxvi,xxvii,xxviii,xxix}. Moreover, the activated Ca²⁺/nitric oxide/cGMP cascade is also closely related to the increased expression of FGF-2 and VEGF, two key regulators of angiogenesis^{xxiii}. In addition, the crosstalk between Ca²⁺, ERK, PKA, and PKG signaling under PEMF stimulation was also reported^{xv,xviii}. All these findings show the prominent role of Ca²⁺ signaling in PEMFs-induced bone repair.

Wnt/β-catenin signaling pathway

Extracellular Wnt ligands bind to their seven-pass transmembrane Frizzled receptors simultaneously with a co-receptor of the arrow/Lrp family (e.g., LRP5 and LRP6), thus stabilizes β-catenin in the cytoplasm and initiates the canonical Wnt/β-catenin signaling pathway^{xxx}. This signaling pathway is conserved throughout metazoans and is essential for cell proliferation, differentiation, development, self-renewal, and cell fate determination^{xxxi,xxxii}.

Much evidence has suggested that the Wnt/β-catenin signaling pathway acts as a key regulator in PEMF-induced osteogenic differentiation of mesenchymal progenitor cells, bone formation and repair. For instance, in vitro assay studies, gene and protein expressions of canonical Wnt/β-catenin signaling pathway, including Wnt1, LRP6, and β-catenin, were all significantly enhanced after PEMF exposure at both proliferation and differentiation stages of osteoblast-like MC3T3-E1 cells^{xxxiii}. In addition, except the upregulation of mRNA expressions of Wnt1, Wnt3a, LRP5 and β-catenin in tissue derived mesenchymal stem cells (ADSCs), PEMFs intervention could also reduce the expression of dickkopf1 (DKK1) which usually acts as an inhibitor of Wnt signaling pathway^{xxxiv}. Furthermore, the enhanced Wnt/β-catenin signaling induced by PEMFs notably elevated the expression of proliferation phase related target genes,

Ccnd 1 and Ccne 1, and differentiation phase related genes, ALP, OCN, COL1, and Runx2, in osteoblast cells, which accelerated the osteoblasts proliferation, differentiation, and mineralization, three pivotal processes of bone formation^{xxvii,xxviii}. On the other hand, according to in vivo assay studies, PEMFs effectively reversed the bone mass loss and deterioration of bone microarchitecture analyzed by microCT and attenuated biomechanical strength deterioration evaluated by three-point bending test in hind limb-suspended ovariectomized rats through the Wnt/Lrp5/ β -catenin signal pathway^{xxxv,xxxvi}, indicating that activating this pathway by PEMF exposure is beneficial for bone disorders^{xxxvii}.

MAPK pathway

The MAPK pathway is important in the transduction of extracellular signals to various cellular compartments and is involved in cell proliferation, differentiation, migration, and death^{xxxviii}. Conventional MAPKs include Erk1/2, JNK, and p38. The MAPK pathway plays a critical role in PEMF-induced osteogenic differentiation and osteoblasts' viability and function. For example, extremely low-frequency pulsed electromagnetic field (ELF-PEMF) treatment could significantly increase the total protein content, mitochondrial activity, and ALP activity and enhance the formation of mineralized matrix of human osteoblasts with a poor initial osteoblast function through triggering the ERK1/2 signaling pathway. When the cells were treated with U0126, an inhibitor of the ERK1/2 signaling cascade, the positive effects of the ELF-PEMF treatment on osteoblast function were abolished^{xxxix}. Other studies also revealed that the MEK/ERK signaling pathway regulated the promoting effects of PEMF on bone marrow mesenchymal stem cell (BMSC) proliferation, expression of osteogenic genes (RUNX2, BSP, OPN), ALP activity, and calcium deposition^{xviii,xxviii,xl,xli}. Additionally, one study reported that the p38 MAPK pathway is involved in the increased production of collagen synthesis in osteoblast-like cells stimulated by ELF-EMF exposure^{xlii}. Interestingly, a recent research suggested that a 45 Hz EMF promoted the osteogenic differentiation of adipose-derived stem cells, whereas a 7.5 Hz EMF directly augmented the expression of osteoclastogenic markers and regulated the osteoclast differentiation through ERK and p38 MAPK activation^{xliii}. This finding indicated that PEMFs can simultaneously influence osteoblastic and osteoclastic activities under defined electromagnetic conditions.

FGF and VEGF pathways

Osteogenesis and angiogenesis, including cell–cell communication between blood vessel cells and bone cells, are essential for bone repair. Many studies suggested that PEMFs play a promotion effect not only in osteogenesis but also in angiogenesis^{xliv,xlv,xlvi,xlvii}. PEMFs may facilitate bone repair by augmenting the interaction between osteogenesis and blood vessel growth. During this complex process, FGF and VEGF, two key angiogenesis-related cytokines, may play critical regulatory roles. The FGF signaling pathway has been demonstrated to contribute in the regulation of proliferation and differentiation of osteoblasts and

in angiogenesis^{xlvii} and the VEGF signaling pathway has also been reported to be involved in a reciprocal, functional, and regulatory relationship between osteoblasts and endothelial cells during osteogenesis^{xlix,l,li}. A study indicated that a 150% increase in FGF-2 mRNA and a fivefold elevation of FGF-2 proteins in human umbilical vein endothelial cells (HUVECs) exposed to PEMF were monitored and the release of functional FGF-2 from PEMF-stimulated HUVECs specially increased endothelial cell proliferation and tubulization, processes that are important for vessel formation^{lii}. KDR/Flk-1, a tyrosine kinase receptor of VEGF, is autophosphorylated in response to VEGF stimulation and is capable of transducing VEGF signals. One research has revealed that PEMF stimulation significantly increased the expression and phosphorylated levels of KDR/Flk-1 and promoted proliferation, migration, and tube formation of HUVECs^{xxxix}. The pro angiogenesis effect through the FGF and VEGF signaling pathways of PEMFs provide another explanation for the therapeutic function of PEMFs in bone repair. Many studies are still required to further clarify the efficacy of FGF and VEGF in PEMF-induced bone repair.

TGF- β /BMP pathway

TGF- β s and BMPs, as multifunctional growth factors, belong to the TGF- β super family. The interaction of TGF- β s/BMPs with TGF- β specific type 1 and type 2 or BMP serine/threonine kinase receptors initiates the signaling cascade via canonical (or Smad-dependent pathways) and non-canonical pathways (or Smad-independent signaling pathways)^{liii}. The TGF- β /BMP signaling pathway plays an important regulatory role in bone repair^{liv,lv,lvi,lvii,lviii,lix}. It is also confirmed to be involved in PEMF-induced osteogenesis. Several studies demonstrated that PEMF stimulation could significantly increase the expression of TGF- β in both osteoblast-like cells and cells from atrophic or hypertrophic non-unions^{xiii,lx,lxi,lxii,lxiii}. Moreover, a recent research suggested that PEMFs activated the TGF- β signaling via Smad2 in differentiated and mineralizing osteoblasts and augmented the expression of osteoblast differentiation marker genes, such as ALP and type I collagen, and exerted its osteogenesis promotional function. The expression of BMPs in osteogenesis was also enhanced by PEMFs according to in vitro and clinical studies^{lxv,lxvi,lxvii}. Furthermore, another recent study revealed that PEMFs stimulate osteogenic differentiation and maturation of osteoblasts by primary cilium-mediated upregulated expression of BMPRII, one of the receptors of BMPs, and subsequently activation of BMP–Smad1/5/8 signaling^{lxviii}. Given the separate promotional effects on the differentiation and maturation of osteoblasts of BMPs and PEMFs, many studies found that combined BMP and PEMF stimulation would augment bone formation to a greater degree than treatment with either stimulus^{lxix,lxx,lxxi,lxxii}.

Other pathways

IGF signaling pathway is also an important signaling implicating in osteoblast differentiation and bone formation^{[lxxiii](#),[lxxiv](#)}. It was reported that PEMFs significantly increase the level of mRNA expression of IGF-1 and promote bone formation in rat femoral tissues in vitro^{[lxxv](#)}. In addition, IGF-1 in combination with PEMFs augmented cartilage explant anabolic activities, increased PG synthesis, restricted the catabolic effect of IL-1b, and showed a synergistic chondroprotective effect on human articular cartilage^{[lxxvi](#)}. Another study showed that dexamethasone combined with PEMF upregulated the mRNA expression of IGF-1 and improved dexamethasone-induced bone loss and osteoporosis^{[lxxvii](#)}. Notch signaling is a highly conserved pathway that regulates cell fate decisions and skeletal development. A recent research advocated that the expression levels of Notch receptor (Notch4) and its ligand DLL4 and nuclear target genes (Hey1, Hes1, and Hes5) were upregulated during the PEMF-induced osteogenic differentiation of hMSCs. Moreover, the Notch pathway inhibitors effectively inhibited the expression of osteogenic markers, including Runx2, Dlx5, Osterix, as well as Hes1 and Hes5, indicating that the Notch signaling plays an important regulatory role in PEMF-induced osteogenic differentiation of hMSCs^{[lxxviii](#)}. The cAMP/PKA signaling pathway is another signaling involved in the PEMF-induced bone repair. Recent studies have demonstrated that PEMFs notably increased the cAMP level and PKA activity and accelerated the osteogenic differentiation of MSCs^{[xxvii](#),[xxxiv](#),[lxxix](#)}.

Therapeutic applications of PEMF in bone repair

The promotional effects of PEMFs on osteogenesis and angiogenesis in bone repair have been well established in either vitro or in vivo animal studies. Several key-signaling pathways involved in PEMF-induced bone repair were elaborate above. Moreover, several decades of PEMF applications in the treatment of skeletal diseases have clearly proved its potential benefit in augmenting bone repair.

Osteoporosis

Osteoporosis is a worldwide health problem with high morbidity, especially in postmenopausal women^{[lxxx](#),[lxxxi](#),[lxxxii](#)}. It is generally defined as a systemic skeletal disease characterized by low bone mineral density (BMD) and compromised bone strength, leading to enhanced bone fragility, increased fracture risk, and resultant disability, which strikingly affects patients' quality of life^{[lxxxiii](#),[lxxxiv](#)}. As PEMFs were verified to be equally effective with mechanical stimulation in maintaining or improving bone mass according to experiments of NASA between 1976 and 1979, many clinical studies have gradually achieved positive therapeutic effects for osteoporosis by PEMF exposure^{[lxxxv](#),[lxxxvi](#),[lxxxvii](#),[lxxxviii](#),[lxxxix](#),[xc](#),[xci](#)}. Chronic pain is a common symptom of

people with osteoporosis. Many randomized controlled trials indicated that PEMF exposure could relieve chronic pain caused by osteoporosis^{lxxxii,lxxxiii}. Moreover, in a study of 126 patients with primary osteoporosis, PEMF provided a faster and significant effect in relieving pain for patients with type I osteoporosis than those with type II⁸⁴. BMD is the gold standard for diagnosing osteoporosis and the best quantitative indicator for forecasting the risk of osteoporotic fracture, monitoring the natural course of osteoporosis, and evaluating the effect of osteoporosis. Tabrah indicated that BMD of the treated radii was elevated notably in the sixth week in a clinical study of 20 women with PMOP treated with PEMFs^{lxxvi}. In Garland's research, which evaluated the effect of PEMFs on knee osteoporosis in individuals with spinal cord injury, BMD was also elevated. At three months, BMD was increased by 5.1% in the stimulated knees but declined to 6.6% in the control knees^{lxxvii}. PEMFs as a noninvasive physical therapy method avoids the defects of pharmacotherapy for osteoporosis, including the multiple side effects, the more cost and the low persistence. More importantly, a randomized, active-controlled clinical trial on postmenopausal osteoporosis (PMO) in Southwest China revealed that PEMFs had the same effect as alendronate, which is, currently, the most commonly prescribed medication for treating PMO within 24 weeks^{xciii}. Furthermore, the hemorheological safety of PEMFs for treating osteoporosis was also observed by a randomized, placebo-controlled clinical study^{xciv}. All these results support the efficiency and safety of PEMFs for osteoporosis treatment and as an advantageous treatment strategy in the future.

Fractures, delayed unions, and non-unions

Fractures, particularly those that had developed into delayed unions or even non-unions, have a substantial clinical, economic, and quality of life impact. Apart from traditional surgical management and rigid fixation (either internal or external), noninvasive PEMFs have already been used effectively in clinics as physical therapy to accelerate and finalize the healing process of a fresh fracture and reactivate the healing process of delayed unions and non-unions for nearly forty years since they were first approved by the US Food and Drug Administration^{iv,v}. A recent systematic review and meta-analysis of randomized controlled trials showed that PEMFs significantly shortened the time to radiological union for acute fractures undergoing non-operative treatment and acute fractures of the upper limb and accelerated the time to clinical union for acute diaphyseal fractures^{xcvi}. Moreover, a prospective study that evaluated the treatment effect of PEMFs on 64 patients undergoing hindfoot arthrodesis (144 joints) revealed that the adjunctive use of a PEMF in elective hindfoot arthrodesis may increase the rate and speed of radiographic union of these joints^{xcvii}. Despite the relative scarcity of well-organized randomized controlled trials, many studies highlight the practice usefulness of PEMFs in treating tibial delayed unions or non-unions, with efficacy up to 87%^{vi,lxxv,xcviii,xcix}. Furthermore, in a broad literature review comparing PEMF treatment of non-unions with surgical therapy, Gossling noted that 81% of reported cases healed with PEMF versus 82% with surgery. Obvious therapeutic advantages of PEMFs were showed compared with surgery in treatment for infected non-unions (81% versus 69%)^c and closed injury caused non-unions (85% versus 79%). In addition, a recent double-blind randomized study advocated that the adjunctive use of PEMF for fifth metatarsal fracture non-unions significantly shortened the average time to complete radiographic union from 14.7 weeks to 8.9 weeks compared with

the control group without PEMF exposure; the elevated expression levels of PIGF, BMP-5, and BMP-7, key regulators of angiogenesis and osteogenesis, were first detected in the non-union environment before and after the application of PEMFs^{lviii}. These studies strikingly support PEMFs as an optional and effective method to accelerate fracture healing.

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|------|--|
| i | Gorissen BM, Wolschrijn CF, van Vilsteren AA, et al. 2016. Trabecular bone of pre-cials at birth; Are they prepared to run for the wolf(f)? J Morphol 2016; 277:948-956. |
| ii | The classic: Fundamental aspects of fracture treatment by Iwao Yasuda , reprinted from J. Kyoto Med. Soc., 4: 395-406, 1953. Clin Orthop Relat Res 1977 5-8. |
| iii | Bassett CA, Mitchell SN, Norton L, et al. 1978. Repair of non-unions by pulsing electromagnetic fields. Acta Orthop Belg 1978; 44:706-724. |
| iv | Gupta AK, Srivastava KP, Avasthi S. 2009. Pulsed electromagnetic stimulation in nonunion of tibial diaphyseal fractures. Indian J Orthop 2009; 43:156-160. |
| v | Meskens MW, Stuyck JA, Feys H, et al. 1990. Treatment of nonunion using pulsed electromagnetic fields: a retrospective follow-up study. Acta Orthop Belg 1990; 56:483-488. |
| vi | Assiotis A, Sachinis NP, Chalidis BE. 2012. Pulsed electromagnetic fields for the treatment of tibial delayed unions and nonunions. A prospective clinical study and review of the literature. J Orthop Surg Res 2012; 7:24. |
| vii | Chalidis B, Sachinis N, Assiotis A, et al. 2011. Stimulation of bone formation and fracture healing with pulsed electromagnetic fields: biologic responses and clinical implications. Int J Immunopathol Pharmacol 2011; 24:17-20. |
| viii | Kuzyk PR, Schemitsch EH. 2009. The science of electrical stimulation therapy for fracture healing. Indian J Orthop 2009; 43:127-131. |
| ix | Wang YY, et al., 2018. Pulsed electromagnetic fields promote bone formation by activating the sAC-cAMP-PKA-CREB signaling pathway. J Cell Physiol. 2018 Aug 1. doi: 10.1002/jcp.27098. |
| x | Cai J, Li W, Sun T, et al. 2018. Pulsed electromagnetic fields preserve bone architecture and mechanical properties and stimulate porous implant osseointegration by promoting bone anabolism in type 1 diabetic rabbits. Osteoporos Int. 2018 May;29(5):1177-1191. |
| xi | Li S, et al., 2018. Magnetic Resonance Spectroscopy for Evaluating the Effect of Pulsed Electromagnetic Fields on Marrow Adiposity in Postmenopausal Women With Osteopenia. J Comput Assist Tomogr. 2018 Sep/Oct;42(5):792-797. |
| xii | Tabrah F, Bassett CA, et al., 1990. Bone density changes in osteoporosis-prone women exposed to pulsed electromagnetic fields (PEMFs). J Bone Miner Res. 1990 May;5(5):437-42. |
| xiii | Lei T, Liang Z, Li F, et al. 2018. Pulsed electromagnetic fields (PEMF) attenuate changes in vertebral bone mass, architecture and strength in ovariectomized mice. Bone. 2018 Mar;108:10-19. |

| | |
|-------|--|
| xiv | Jiang Y, Gou H, Wang S, et al. 2016. Effect of Pulsed Electromagnetic Field on Bone Formation and Lipid Metabolism of Glucocorticoid-Induced Osteoporosis Rats through Canonical Wnt Signaling Pathway. <i>Evid Based Complement Alternat Med.</i> 2016;2016:4927035. |
| xv | He Z, Selvamurugan N, et al. 2018. Pulsed electromagnetic fields inhibit human osteoclast formation and gene expression via osteoblasts. <i>Bone.</i> 2018 Jan;106:194-203. |
| xvi | Wu S, Yu Q, et al. 2018. Synergistic effect of a LPEMF and SPIONs on BMMSC proliferation, directional migration, and osteoblastogenesis. <i>Am J Transl Res.</i> 2018 May 15;10(5):1431-1443. |
| xvii | Li J, Zeng Z, Zhao, Y et al. 2017. Effects of low-intensity pulsed electromagnetic fields on bone microarchitecture, mechanical strength and bone turnover in type 2 diabetic db/db mice. <i>Sci Rep.</i> 2017 Sep 7;7(1):10834. |
| xviii | Li B, Bi J, Li W. 2017. Effects of pulsed electromagnetic fields on histomorphometry and osteocalcin in disuse osteoporosis rats. <i>Technol Health Care.</i> 2017 Jul 20;25(S1):13-20. |
| xix | Jing D, Zhai M, Tong S et al. 2016. Pulsed electromagnetic fields promote osteogenesis and osseointegration of porous titanium implants in bone defect repair through a Wnt/ β -catenin signaling-associated mechanism. <i>Sci Rep.</i> 2016 Aug 24;6:32045. |
| xx | Yuan J, Xin F, Jiang W. 2018. Underlying Signaling Pathways and Therapeutic Applications of Pulsed Electromagnetic Fields in Bone Repair. <i>Cell Physiol Biochem.</i> 2018;46(4):1581-1594. |
| xxi | Li JK, Lin JC, Liu HC, et al. 2006. Comparison of ultrasound and electromagnetic field effects on osteoblast growth. <i>Ultrasound Med Biol</i> 2006; 32: 769-775. |
| xxii | Pall ML. 2013. Electromagnetic fields act via activation of voltage-gated calcium channels to produce beneficial or adverse effects. <i>J Cell Mol Med</i> 2013; 17: 958-965. |
| xxiii | Petecchia L, Sbrana F, Utzeri R, et al. 2015. Electro-magnetic field promotes osteogenic differentiation of BM-hMSCs through a selective action on Ca(2+)-related mechanisms. <i>Sci Rep</i> 2015; 5: 13856. |
| xxiv | Kim MO, Jung H, Kim SC, et al. 2015. Electromagnetic fields and nanomagnetic particles increase the osteogenic differentiation of human bone marrow-derived mesenchymal stem cells. <i>Int J Mol Med</i> 2015; 35: 153-160. |
| xxv | Zhong C, Zhao TF, Xu ZJ, et al. 2002. Effects of electromagnetic fields on bone regeneration in experimental and clinical studies: a review of the literature. <i>Chin Med J (Engl)</i> 2012; 125: 367-372. |
| xxvi | Diniz P, Soejima K, Ito G. 2002. Nitric oxide mediates the effects of pulsed electromagnetic field stimulation on the osteoblast proliferation and differentiation. <i>Nitric Oxide</i> 2002; 7: 18-23. |
| xxvii | Cheng G, Zhai Y, Chen K, et al. 2011. Sinusoidal electromagnetic field stimulates rat osteoblast differentiation and maturation via activation of NO-cGMP-PKG pathway. <i>Nitric Oxide</i> 2011; 25: 316-325. |

| | |
|---------|---|
| xxviii | Pilla A, Fitzsimmons R, Muehsam D, et al. 2011. Electromagnetic fields as first messenger in biological signaling: Application to calmodulin-dependent signaling in tissue repair. <i>Biochim Biophys Acta</i> 2011; 1810: 1236-1245. |
| xxix | Nelson FR, Zvirbulis R, Pilla AA. 2013. Non-invasive electromagnetic field therapy produces rapid and substantial pain reduction in early knee osteoarthritis: a randomized double-blind pilot study. <i>Rheumatol Int</i> 2013; 33: 2169-2173. |
| xxx | Drenser KA. 2016. Wnt signaling pathway in retinal vascularization. <i>Eye Brain</i> 2016;8:141-146. |
| xxxi | Ramakrishnan AB, Cadigan KM. 2017. Wnt target genes and where to find them. <i>F1000Res</i> 2017;6:746. |
| xxxii | Pai SG, Carneiro BA, Mota JM, et al. 2017. Wnt/beta-catenin pathway: modulating anticancer immune response. <i>J Hematol Oncol</i> 2017;10:101. |
| xxxiii | Zhai M, Jing D, Tong S, et al. 2016. Pulsed electromagnetic fields promote in vitro osteoblastogenesis through a Wnt/beta-catenin signaling-associated mechanism. <i>Bioelectromagnetics</i> 2016; 10.1002/bem.21961. |
| xxxiv | Fathi E, Farahzadi R. 2017. Enhancement of osteogenic differentiation of rat adipose tissue-derived mesenchymal stem cells by zinc sulphate under electromagnetic field via the PKA, ERK1/2 and Wnt/beta-catenin signaling pathways. <i>PLoS One</i> 2017; 12:e0173877. |
| xxxv | Jing D, Cai J, Wu Y, et al. 2014. Pulsed electromagnetic fields partially preserve bone mass, microarchitecture, and strength by promoting bone formation in hindlimb-suspended rats. <i>J Bone Miner Res</i> 2014;29:2250-2261. |
| xxxvi | Jing D, Li F, Jiang M, et al. 2013. Pulsed electromagnetic fields improve bone microstructure and strength in ovariectomized rats through a Wnt/Lrp5/beta-catenin signaling-associated mechanism. <i>PLoS One</i> 2013; 8:e79377. |
| xxxvii | Wu S, Yu Q, et al., 2018. Pulsed electromagnetic field induces Ca ²⁺ -dependent osteoblasto-genesis in C3H10T1/2 mesenchymal cells through the Wnt-Ca ²⁺ /Wnt-β-catenin signaling pathway. <i>Biochem Biophys Res Commun.</i> 2018 Sep 5;503(2):715-721 |
| xxxviii | Lake D, Correa SA, Muller J. 2016. Negative feedback regulation of the ERK1/2 MAPK pathway. <i>Cell Mol Life Sci</i> 2016; 73:4397-4413. |
| xxxix | Ehnert S, Falldorf K, Fentz AK, et al. 2015. Primary human osteoblasts with reduced alkaline phosphatase and matrix mineralization baseline capacity are responsive to extremely low frequency pulsed electromagnetic field exposure-Clinical implication possible. <i>Bone Rep</i> 2015; 3:48-56. |
| xl | Song MY, Yu JZ, Zhao DM, et al. 2014. The time-dependent manner of sinusoidal electromagnetic fields on rat bone marrow mesenchymal stem cells proliferation, differentiation, and mineralization. <i>Cell Biochem Biophys</i> 2014;69:47-54. |
| xli | Yong Y, Ming ZD, Feng L, et al. 2016. Electromagnetic fields promote osteogenesis of rat mesenchymal stem cells through the PKA and ERK1/2 pathways. <i>J Tissue Eng Regen Med</i> 2016;10:E537-E545. |

| | |
|--------|---|
| xlii | Soda A, Ikehara T, Kinouchi Y, et al. 2008. Effect of exposure to an extremely low frequency-electromagnetic field on the cellular collagen with respect to signaling pathways in osteoblast-like cells. <i>J Med Invest</i> 2008; 55:267-278. |
| xliii | Hong JM, Kang KS, Yi HG, et al. 2014. Electromagnetically controllable osteoclast activity. <i>Bone</i> 2014;62:99-107. |
| xliv | Yen-Patton GP, Patton WF, et al. 1988. Endothelial cell response to pulsed electromagnetic fields: stimulation of growth rate and angiogenesis in vitro. <i>J Cell Physiol</i> 1988; 134:37-46. |
| xlv | Hopper RA, VerHalen JP, Tepper O, et al. 2009. Osteoblasts stimulated with pulsed electromagnetic fields increase HUVEC proliferation via a VEGF-A independent mechanism. <i>Bioelectromagnetics</i> 2009; 30:189-197. |
| xlvi | Delle Monache S, Alessandro R, Iorio R, et al. 2008. Extremely low frequency electromagnetic fields (ELF-EMFs) induce in vitro angiogenesis process in human endothelial cells. <i>Bioelectromagnetics</i> 2008; 29:640-648. |
| xlvii | Callaghan MJ, Chang EI, Seiser N, et al. 2008. Pulsed electromagnetic fields accelerate normal and diabetic wound healing by increasing endogenous FGF-2 release. <i>Plast Reconstr Surg</i> 2008; 121:130-141. |
| xlviii | Yun YR, Won JE, Jeon E, et al. 2010. Fibroblast growth factors: biology, function, and application for tissue regeneration. <i>J Tissue Eng</i> 2010; 2010:218142. |
| xl ix | Deckers MM, Karperien M, van der Bent C, et al. 2000. Expression of vascular endothelial growth factors and their receptors during osteoblast differentiation. <i>Endocrinology</i> 2000; 141:1667-1674. |
| l | Deckers MM, van Bezooijen RL, van der Horst G, et al. 2002. Bone morphogenetic proteins stimulate angiogenesis through osteoblast-derived vascular endothelial growth factor A. <i>Endocrinology</i> 2002; 143:1545-1553. |
| li | Villars F, Bordenave L, Bareille R, et al. 2000. Effect of human endothelial cells on human bone marrow stromal cell phenotype: role of VEGF? <i>J Cell Biochem</i> 2000; 79:672-685. |
| lii | Tepper OM, Callaghan MJ, Chang EI, et al. 2004. Electromagnetic fields increase in vitro and in vivo angiogenesis through endothelial release of FGF-2. <i>FASEB J</i> 2004; 18:1231-1233. |
| liii | Carreira AC, Lojudice FH, Halcsik E, et al. 2014. Bone morphogenetic proteins: facts, challenges, and future perspectives. <i>J Dent Res</i> 2014; 93:335-345. |
| liv | Gao Y, Zhang Y, Lu Y, et al. 2016. TOB1 Deficiency Enhances the Effect of Bone Marrow-Derived Mesenchymal Stem Cells on Tendon-Bone Healing in a Rat Rotator Cuff Repair Model. <i>Cell Physiol Biochem</i> 2016; 38:319-329. |
| lv | Liao J, Wei Q, Zou Y, et al. 2017. Notch Signaling Augments BMP9-Induced Bone Formation by Promoting the Osteogenesis-Angiogenesis Coupling Process in Mesenchymal Stem Cells (MSCs). <i>Cell Physiol Biochem</i> 2017; 41:1905-1923. |
| lvi | Peng WX, Wang L. 2017. Adenovirus-Mediated Expression of BMP-2 and BFGF in Bone Marrow Mesenchymal Stem Cells Combined with Demineralized Bone Matrix For Repair of Femoral Head Osteonecrosis in Beagle Dogs. <i>Cell Physiol Biochem</i> 2017; 43:1648-1662. |

| | |
|--------|--|
| lvii | Wang R, Xu B, Xu HG. 2017. Up-Regulation of TGF-beta Promotes Tendon-to-Bone Healing after Anterior Cruciate Ligament Reconstruction using Bone Marrow-Derived Mesenchymal Stem Cells through the TGF-beta/ MAPK Signaling Pathway in a New Zealand White Rabbit Model. <i>Cell Physiol Biochem</i> 2017; 41:213-226. |
| lviii | Zhou W, Yu L, Fan J, et al. 2017. Endogenous Parathyroid Hormone Promotes Fracture Healing by Increasing Expression of BMPR2 through cAMP/PKA/CREB Pathway in Mice. <i>Cell Physiol Biochem</i> 2017; 42:551-563. |
| lix | Zou L, Zhang G, Liu L, et al. 2017. A MicroRNA-124 Polymorphism is Associated with Fracture Healing via Modulating BMP6 Expression. <i>Cell Physiol Biochem</i> 2017; 41:2161-2170. |
| lx | Guerkov HH, Lohmann CH, Liu Y, et al. 2001. Pulsed electromagnetic fields increase growth factor release by nonunion cells. <i>Clin Orthop Relat Res</i> 2001;265-279. |
| lxi | Kang KS, Hong JM, Seol YJ, et al. 2015. Short-term evaluation of electromagnetic field pretreatment of adipose-derived stem cells to improve bone healing. <i>J Tissue Eng Regen Med</i> 2015; 9:1161-1171. |
| lxii | Ding S, Peng H, Fang HS, et al. 2011. Pulsed electromagnetic fields stimulation prevents steroid-induced osteonecrosis in rats. <i>BMC Musculoskelet Disord</i> 2011; 12:215. |
| lxiii | Lohmann CH, Schwartz Z, Liu Y, et al. 2000. Pulsed electromagnetic field stimulation of MG63 osteoblast-like cells affects differentiation and local factor production. <i>J Orthop Res</i> 2000; 18:637-646. |
| lxiv | Selvamurugan N, He Z, Rifkin D, et al. 2017. Pulsed Electromagnetic Field Regulates MicroRNA 21 Expression to Activate TGF-beta Signaling in Human Bone Marrow Stromal Cells to Enhance Osteoblast Differentiation. <i>Stem Cells Int</i> 2017; 2017:2450327. |
| lxv | Streit A, Watson BC, Granata JD, et al. 2016. Effect on Clinical Outcome and Growth Factor Synthesis With Adjunctive Use of Pulsed Electromagnetic Fields for Fifth Metatarsal Nonunion Fracture: A Double-Blind Randomized Study. <i>Foot Ankle Int</i> 2016; 37:919-923. |
| lxvi | Bodamyali T, Bhatt B, Hughes FJ, et al. 1998. Pulsed electromagnetic fields simultaneously induce osteogenesis and upregulate transcription of bone morphogenetic proteins 2 and 4 in rat osteoblasts in vitro. <i>Biochem Biophys Res Commun</i> 1998; 250: 458-461. |
| lxvii | Zhou J, Ming LG, Ge BF, et al. 2011. Effects of 50 Hz sinusoidal electromagnetic fields of different intensities on proliferation, differentiation and mineralization potentials of rat osteoblasts. <i>Bone</i> 2011; 49:753-761 |
| lxviii | Xie YF, Shi WG, Zhou J, et al. 2016. Pulsed electromagnetic fields stimulate osteogenic differentiation and maturation of osteoblasts by upregulating the expression of BMPRII localized at the base of primary cilium. <i>Bone</i> 2016; 93:22-32. |
| lxix | Selvamurugan N, Kwok S, Vasilov A, et al. 2007. Effects of BMP-2 and pulsed electromagnetic field (PEMF) on rat primary osteoblastic cell proliferation and gene expression. <i>J Orthop Res</i> 2007; 25:1213-1220. |

| | |
|----------------|--|
| lxx | Schwartz Z, Simon BJ, Duran MA, et al. 2008. Pulsed electromagnetic fields enhance BMP-2 dependent osteoblastic differentiation of human mesenchymal stem cells. <i>J Orthop Res</i> 2008; 26:1250-1255. |
| lxxi | Ongaro A, Pellati A, Bagheri L, et al. 2014. Pulsed electromagnetic fields stimulate osteogenic differentiation in human bone marrow and adipose tissue derived mesenchymal stem cells. <i>Bioelectromagnetics</i> 2014; 35:426-436. |
| lxxii | Yang HJ, Kim RY, Hwang SJ. 2015. Pulsed Electromagnetic Fields Enhance Bone Morphogenetic Protein-2 Dependent-Bone Regeneration. <i>Tissue Eng Part A</i> 2015; 21:2629-2637. |
| lxxiii | Arvidson K, Abdallah BM, Applegate LA, et al. 2011. Bone regeneration and stem cells. <i>J Cell Mol Med</i> 2011; 15:718-746. |
| lxxiv | Guo Y, Tang CY, Man XF, et al. 2017. Insulin-like growth factor-1 promotes osteogenic differentiation and collagen I alpha 2 synthesis via induction of mRNA-binding protein LARP6 expression. <i>Dev Growth Differ</i> 2017; 59:94-103. |
| lxxv | Zhou J, Ma XN, Gao YH, et al. 2016. Sinusoidal electromagnetic fields promote bone formation and inhibit bone resorption in rat femoral tissues in vitro. <i>Electromagn Biol Med</i> 2016; 35:75-83. |
| lxxvi | Ongaro A, Pellati A, Masieri FF, et al. 2011. Chondroprotective effects of pulsed electromagnetic fields on human cartilage explants. <i>Bioelectromagnetics</i> 2011; 32:543-551. |
| lxxvii | Esmail MY, Sun L, Yu L, et al. 2012. Effects of PEMF and glucocorticoids on proliferation and differentiation of osteoblasts. <i>Electromagn Biol Med</i> 2012; 31:375-381. |
| lxxviii | Bagheri L, Pellati A, Rizzo P, et al. 2017. Notch pathway is active during osteogenic differentiation of human bone marrow mesenchymal stem cells induced by pulsed electromagnetic fields. <i>J Tissue Eng Regen Med</i> 2017; 10.1002/term.2455 |
| lxxix | Fang QQ, Li ZZ, Zhou J, et al. 2016. Low-frequency pulsed electromagnetic fields promotes rat osteoblast differentiation in vitro through cAMP/PKA signal pathway. <i>Nan Fang Yi Ke Da Xue Xue Bao</i> 2016; 36:1508-1513. |
| lxxx | Pai MV. 2017. Osteoporosis Prevention and Management. <i>J Obstet Gynaecol India</i> 2017; 67:237-242. |
| lxxxi | Golob AL, Laya MB. 2015. Osteoporosis: screening, prevention, and management. <i>Med Clin North Am</i> 2015; 99:587-606. |
| lxxxii | Verbovoy AF, Pashentseva AV, Sharonova LA. 2017. Osteoporosis: Current state of the art. <i>Ter Arkh</i> 2017; 89:90-97. |
| lxxxiii | Ensrud KE, Crandall CJ. 2017. Osteoporosis. <i>Ann Intern Med</i> 2017; 167:ITC17-ITC32. |
| lxxxiv | Watts NB, Bilezikian JP, Camacho PM, et al. 2010. American Association of Clinical Endocrinologists Medical Guidelines for Clinical Practice for the diagnosis and treatment of postmenopausal osteoporosis. <i>Endocr Pract</i> 2010; 16:S1-37. |
| lxxxv | Rubin CT, McLeod KJ, Lanyon LE. 1989. Prevention of osteoporosis by pulsed electromagnetic fields. <i>J Bone Joint Surg Am</i> 1989; 71:411-417. |
| lxxxvi | Tabrah F, Hoffmeier M, Bassett CA, et al. 1990. Bone density changes in osteoporosis-prone women exposed to pulsed electromagnetic fields (PEMFs). <i>J Bone Miner Res</i> 1990; 5:437-442. |

| | |
|----------|--|
| lxxxvii | Garland DE, Adkins RH, Matsuno NN, et al. 1999. The effect of pulsed electromagnetic fields on osteoporosis at the knee in individuals with spinal cord injury. <i>J Spinal Cord Med</i> 1999; 22:239-245. |
| lxxxviii | Eyres KS, Saleh M, Kanis JA. 1996. Effect of pulsed electromagnetic fields on bone formation and bone loss during limb lengthening. <i>Bone</i> 1996; 18:505-509. |
| lxxxix | Liu H, Liu Y, Yang L, et al. 2014. Curative effects of pulsed electromagnetic fields on postmenopausal osteoporosis. <i>Sheng Wu Yi Xue Gong Cheng Xue Za Zhi</i> 2014; 31:48-52. |
| xc | Wang R, Wu H, Yang Y, et al. 2016. Effects of electromagnetic fields on osteoporosis: A systematic literature review. <i>Electromagn Biol Med</i> 2016; 35:384-390. |
| xc | Weng YX, Gao QY, Shao HWy, et al. 2003. Osteoporosis pain and effectiveness of pulsed electromagnetic fields in treating pain in patients with osteoporosis. <i>Chin J Osteoporos (China)</i> 2003; 9:3-17. |
| xcii | Hayashi Y. 2007. Bone diseases with Pain. <i>Osteoporosis. Clin Calcium</i> 2007; 17:606-612. |
| xciii | Liu HF, Yang L, He HC, et al. 2013. Pulsed electromagnetic fields on postmenopausal osteoporosis in Southwest China: a randomized, active-controlled clinical trial. <i>Bioelectromagnetics</i> 2013; 34:323-332. |
| xciv | Liu H, Yang L, He H, et al. 2013. The hemorheological safety of pulsed electromagnetic fields in postmenopausal women with osteoporosis in southwest China: a randomized, placebo controlled clinical trial. <i>Clin Hemorheol Microcirc</i> 2013; 55:285-295. |
| xcv | Victoria G, Petrisor B, Drew B, et al. 2009. Bone stimulation for fracture healing: What's all the fuss? <i>Indian J Orthop</i> 2009; 43:117-120. |
| xcvi | Hannemann PF, Mommers EH, Schots JP, et al. 2014. The effects of low-intensity pulsed ultrasound and pulsed electromagnetic fields bone growth stimulation in acute fractures: a systematic review and meta-analysis of randomized controlled trials. <i>Arch Orthop Trauma Surg</i> 2014; 134:1093-1106. |
| xcvii | Dhawan SK, Conti SF, Towers J, et al. 2004. The effect of pulsed electromagnetic fields on hindfoot arthrodesis: a prospective study. <i>J Foot Ankle Surg</i> 2004; 43:93-96. |
| xcviii | Bassett CA, Mitchell SN, Gaston SR. 1981. Treatment of ununited tibial diaphyseal fractures with pulsing electromagnetic fields. <i>J Bone Joint Surg Am</i> 1981; 63:511-523. |
| xcix | de Haas WG, Watson J, Morrison DM. 1980. Non-invasive treatment of ununited fractures of the tibia using electrical stimulation. <i>J Bone Joint Surg Br</i> 1980; 62-B:465-470 |
| c | Nelson FR, Brighton CT, Ryaby J, et al. 2003. Use of physical forces in bone healing. <i>J Am Acad Orthop Surg</i> 2003; 11:344-354. |



Swiss Bionic Solutions Schweiz GmbH

Schulhausstrasse 17 | 8834 Schindellegi, Schweiz

Phone: +41 (62) 295 5951 | Fax: +41 (62) 295 5952 | E-Mail: ch@swissbionic.com

Swiss Bionic Solutions Deutschland GmbH

Biberacher Str. 87 | 88339 Bad Waldsee, Deutschland

Phone: +49 (7524) 996 950 | Fax: +49 (7524) 996 9518 | E-Mail: de@swissbionic.com

Swiss Bionic Solutions USA Inc.

12330 SW 53rd Street | Suite 703 & 704 | Cooper City | Florida 33330, USA

Phone: +1 (954) 766 4153 | Fax: +1 (954) 766 4156 | E-Mail: us@swissbionic.com

Swiss Bionic Solutions Canada Inc.

1195 North Service Rd W. Unit B8 | Oakville, ON, L6M 2W2, Canada

Phone: +1 (905) 465 0753 | Fax: +1 (1 866) 792 8182 | E-Mail: ca@swissbionic.com

Swiss Bionic Solutions Asia Ltd.

998 Canton Road | Mongkok | Kowloon | Hong Kong

Phone: +852 2337-8774 | E-Mail: asia@swissbionic.com

